DRG® Borrelia IgG VIsE ELISA (EIA-4289)

CE

RUO in the USA

Revised 25 July 2012 rm (Vers. 8.1)

This kit is intended for Research Use Only.

Not for use in diagnostic procedures.

Please use only the valid version of the package insert provided with the kit.

INTENDED USE

Enzyme immunoassay for measurement of IgG antibodies against Borrelia burgdorferi in human serum, plasma and CSF. Infections with all three B. burgdorferi subspecies (garinii, afzelii and sensu strictu) are detected.

TEST PRINCIPLE

Solid phase enzyme-linked immunosorbent assay (ELISA) based on the sandwich principle. The wells are coated with antigen. Specific antibodies of the sample binding to the antigen coated wells are detected by a secondary enzyme conjugated antibody (E-Ab) specific for human IgG. After the substrate reaction the intensity of the developed color is proportional to the amount of detected IgG-specific antibodies. Results of samples can be determined directly using the standard curve or calibrator standard.

WARNINGS AND PRECAUTIONS

1. For professional use only.
2. Before starting the assay, read the instructions completely and carefully. Use the valid version of the package insert provided with the kit. Be sure that everything is understood.
3. In case of severe damage of the kit package please contact DRG or your supplier in written form, latest one week after receiving the kit. Do not use damaged components in test runs, but keep safe for complaint related issues.
4. Obey lot number and expiry date. Do not mix reagents of different lots. Do not use expired reagents.
5. Follow good laboratory practice and safety guidelines. Wear lab coats, disposable latex gloves and protective glasses where necessary.
6. Reagents of this kit containing hazardous material may cause eye and skin irritations. See MATERIALS SUPPLIED and labels for details. Material Safety Data Sheets for this product are available upon request.
7. Chemicals and prepared or used reagents have to be treated as hazardous waste according to national biohazard and safety guidelines or regulations.
8. Avoid contact with Stop solution. It may cause skin irritations and burns.
9. All reagents of this kit containing human serum or plasma have been tested and were found negative for anti-HIV I/II, HBsAg and anti-HCV. However, a presence of these or other infectious agents cannot be excluded absolutely and therefore reagents should be treated as potential biohazards in use and for disposal.

STORAGE AND STABILITY

The kit is shipped at ambient temperature and should be stored at 2°-8°C. Keep away from heat or direct sun light. The storage and stability of specimen and prepared reagents is stated in the corresponding chapters.
The microtiter strips are stable up to 3 months after the first opening when stored at 2°-8°C in the tightly closed bag

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SPECIMEN COLLECTION AND STORAGE

Serum, Plasma (EDTA)

The usual precautions for venipuncture should be observed. It is important to preserve the chemical integrity of a blood specimen from the moment it is collected until it is assayed. Do not use grossly hemolytic, icteric or grossly lipemic specimens. Samples appearing turbid should be centrifuged before testing to remove any particulate material.

Storage Serum/Plasma/CSF:	2 °C - 8 °C	≤ -20 °C (Aliquots)	Keep away from heat or direct sun light. Avoid repeated freeze-thaw cycles.
Stability Serum/Plasma/CSF:	5 days	12 months	

MATERIALS SUPPLIED

Quantity	Symbol	Component
1 x 12 x 8	MTP	**Microtiter Plate** Break apart strips. Coated with specific antigen.
1 x 12 mL	ENZCONJ	**Enzyme Conjugate** Ready to use. Green colored. Contains: anti-human IgG, conjugated to peroxidase.
1 x 4 x 1.5 mL	CAL A-D	**Standard A–D** 2; 10; 50; 200 U/mL Standard B = Calibrator Standard Ready to use. Contains: IgG antibodies against B. burgdorferi, stabilizers.
1 x 1.5 mL	High CONTROL	**High Control** Ready to use. Contains: IgG antibodies against B. burgdorferi, stabilizers.
1 x 1.5 mL	Low CONTROL	**Low Control** Ready to use. Contains: human serum, stabilizers.
1 x 100 mL	DILBUF	**Diluent Buffer** Ready to use. Blue colored.
1 x 100 mL	WASHBUF CONC	**Wash Buffer, Concentrate (10x)** Contains: phosphate buffer.
1 x 12 mL	TMB SUBS	**TMB Substrate Solution** Ready to use. Contains: TMB, Buffer, stabilizers.
1 x 12 mL	TMB STOP	**TMB Stop Solution** Ready to use. 1 M H_2SO_4.

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MATERIALS REQUIRED BUT NOT SUPPLIED

1. Micropipettes (Multipette Eppendorf or similar devices, < 3% CV). Volume: 5; 10; 100; 1000 µL (adjustable)
2. Vortex mixer
3. Tubes (≥ 1 mL) for sample dilution
4. Incubator, 37°C
5. 8-Channel Micropipettor with reagent reservoirs
6. Wash bottle, automated or semi-automated microtiter plate washing system
7. Microtiter plate reader capable of reading absorbance at 450 nm (reference wavelength 600-650 nm)
8. Bidistilled or deionised water
9. Paper towels, pipette tips and timer

PROCEDURE NOTES

1. Any improper handling of samples or modification of the test procedure may influence the results. The indicated pipetting volumes, incubation times, temperatures and pretreatment steps have to be performed strictly according to the instructions. Use calibrated pipettes and devices only.
2. Once the test has been started, all steps should be completed without interruption. Make sure that required reagents, materials and devices are prepared ready at the appropriate time. Allow all reagents and specimens to reach room temperature (18-25°C) and gently swirl each vial of liquid reagent and sample before use. Mix reagents without foaming.
3. Avoid contamination of reagents, pipettes and wells/tubes. Use new disposable plastic pipette tips for each component and specimen. Do not interchange caps. Always cap not used vials. Do not reuse wells/tubes or reagents.
4. It is advised to determine samples in duplicate to be able to identify potential pipetting errors.
5. Use a pipetting scheme to verify an appropriate plate layout.
6. Incubation time affects results. All wells should be handled in the same order and time sequences. It is recommended to use an 8-channel Micropipettor for pipetting of solutions in all wells.
7. Microplate washing is important. Improperly washed wells will give erroneous results. It is recommended to use a multichannel pipette or an automatic microplate washing system. Do not allow the wells to dry between incubations. Do not scratch coated wells during rinsing and aspiration. Rinse and fill all reagents with care. While rinsing, check that all wells are filled precisely with Wash Buffer, and that there are no residues in the wells.
8. Humidity affects the coated wells/tubes. Do not open the pouch until it reaches room temperature. Unused wells/tubes should be returned immediately to the resealed pouch including the desiccant.

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PRE-TEST SETUP INSTRUCTIONS

Preparation of lyophilized or concentrated components

Dilute/ dissolve	Component		Diluent	Relation	Remarks	Storage	Stability
100 mL	WASHBUF CONC	ad 1000 mL	bidist. water	1:10	Resolve crystals at 18 °C – 25 °C.	2 °C - 8 °C	2 months

Dilution of Samples

Serum, Plasma

Sample	to be diluted	with	Relation	Remarks
Serum, Plasma	generally	DILBUF	1:101	e.g. 10 µL + 1 mL

Samples containing concentrations higher than the highest standard have to be diluted further.

Serum/CSF

For measurement of cerebrospinal fluid (CSF) according to Reiber, it is necessary to use approximately similar concentrations or Calibrator indices (COI) in the OD range of 1.0 to 0.1 for serum and CSF. This is generally ensured with the following dilutions:

Sample	to be diluted	with	Relation	Remarks
Serum	generally	DILBUF	1:401	e.g. 5 µL + 2 mL
CSF	generally	DILBUF	1:4	50 µL + 150 µL

The Calibrator indices are corrected by the dilution factors of each dilution in relation to the 1:101 dilution:
The Calibrator index for the 1:401 serum dilution must be multiplied by 4 and the 1:4 CSF dilution must be divided by 25.
A set of dilutions should be performed, if the test sample results are not within the range of 1.0 to 0.1 OD. The following dilutions are recommended:

Serum	1:100	1:200	1:400	1:800	1:1600
CSF	1:2	1:4	1:8	1:16	1:32

TEST PROCEDURE

1. Pipette **100 µL** of **each Standard, Control and diluted sample** into the respective wells of the Microtiter Plate. In the qualitative test only Standard B (Calibrator Standard) is used.
2. **Incubate 1 h** at **37°C**. Use adhesive foil or moisture chamber.
3. Remove adhesive foil. Discard incubation solution. Wash plate **3 x** with **300 µL** of diluted **Wash Buffer**. Remove excess solution by tapping the inverted plate on a paper towel.
4. Pipette **100 µL** of **Enzyme Conjugate** into each well.
5. **Incubate 30 min** at **37°C**. Use adhesive foil or moisture chamber.
6. Remove adhesive foil. Discard incubation solution. Wash plate **3 x** with **300 µL** of diluted **Wash Buffer**. Remove excess solution by tapping the inverted plate on a paper towel.

7. For adding of Substrate and Stop Solution use, if available, an 8-channel Micropipettor. Pipetting should be carried out in the same time intervals for Substrate and Stop Solution. Use positive displacement and avoid formation of air bubbles.
8. Pipette **100 µL** of **TMB Substrate Solution** into each well.
9. **Incubate 30 min** at **RT in the dark.**
10. Stop the substrate reaction by adding **100 µL** of **TMB Stop Solution** into each well. Briefly mix contents by gently shaking the plate.
11. Measure optical density with a photometer at **450 nm** (Reference-wavelength: 600-650 nm) within **60 min** after pipetting of the Stop Solution.

QUALITY CONTROL

The test results are only valid if the test has been performed following the instructions. Moreover the user must strictly adhere to the rules of GLP (Good Laboratory Practice) or other applicable standards/laws. Each laboratory should use known samples as further controls.
It is recommended to participate at appropriate quality assessment trials.
The following technical issues should be proven in case of problems: Expiration dates of (prepared) reagents, storage conditions, pipettes, devices, incubation conditions and washing methods.

CALCULATION OF RESULTS

The evaluation of the test can be performed either qualitatively or quantitatively.

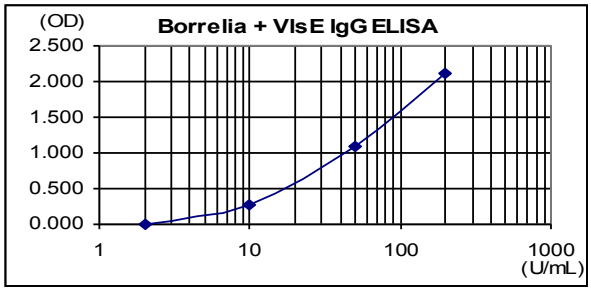
Qualitative Evaluation

The Calibrator value is given by the optical density (OD) of the Standard B (Calibrator standard). The Calibrator index (COI) is calculated from the mean optical densities of the sample and Calibrator value.

Typical Example:
Calibrator = OD (Standard B, Calibrator standard) = 0.45
Sample OD = 0.60
Calibrator index (COI): 0.60 / 0.45 = 1.33. The sample has to be considered high.

Quantitative Evaluation

The obtained OD of the standards (y-axis, linear) are plotted against their concentration (x-axis, logarithmic) either on semi-logarithmic graph paper or using an automated method. A good fit is provided with cubic spline, 4 Parameter Logisitcs or Logit-Log.
For the calculation of the standard curve, apply each signal of the standards (one obvious outlier of duplicates might be omitted and the more plausible single value might be used).
The concentration of the samples can be read from the standard curve.
The initial dilution has been taken into consideration when reading the results from the graph. Results of samples of higher predilution have to be multiplied with the dilution factor.
Samples showing concentrations above the highest standard can be diluted as described in PRE-TEST SETUP INSTRUCTIONS and reassayed.



Typical Calibration Curve
(Example. Do not use for calculation!)

Standard	U/mL	Mean OD
A	2	0.008
B	10	0.267
C	50	1.097
D	200	2.114

LIMITATIONS OF THE PROCEDURE

Specimen collection has a significant effect on the test results. See SPECIMEN COLLECTION AND STORAGE for details.
For cross-reactivities, see PERFORMANCE.
Azide and thimerosal at concentrations > 0.1 % interfere in this assay and may lead to false results.
The following blood components do not have a significant effect (+/- 20 %of expected) on the test results up to the below stated concentrations:

Hemoglobin	2.0 mg/mL
Bilirubin	0.3 mg/mL
Triglyceride	2.5 mg/mL

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PRODUCT REFERENCES

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