TruQuick[™] CRP 10T

A semiquantitative rapid test to detect CRP in whole blood, serum or plasma.

REF TQ4210	IVD	Rx Only
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INTENDED LISE

TruQuick CRP is a rapid chromatographic immunoassay for the semiguantitative detection of human CRP in whole blood, serum or plasma as an aid in the diagnosis of inflammatory conditions.

SUMMARY AND EXPLANATION OF THE TEST

C-reactive Protein (CRP) in patient's sera has been found in association with acute infections, necrotic conditions and a variety of inflammatory disorders. There is a strong correlation between serum levels of CRP and the onset of the inflammatory process. Monitoring the levels of CRP in patient's sera indicates the effectiveness of treatment and the assessment of patient recovery. It is used in particular to differentiate bacterial infections from viral infections

BIOLOGICAL PRINCIPLES

TruQuick CRP detects C-reactive Protein through visual interpretation of color development on the internal strip. Anti-CRP antibodies are immobilized on the test region of the membrane. During testing, the specimen reacts with anti-CRP antibodies conjugated to colored particles and precoated on the sample pad of the test. The mixture then migrates through the membrane by capillary action, and interacts with reagents on the membrane. If the intensity of the test band is weaker than reference band (R), it indicates that the CRP level in the specimen is between 10-30 mg/L. If the intensity of the test band (T) is stronger than the reference band (R), it indicates that the CRP level is above 30 mg/L. The appearance of control line serves as a procedural control, indicating that the proper volume of specimen has been added and membrane wicking has occurred.

REAGENTS/MATERIALS PROVIDED

The maximum number of tests obtained from this test kit is listed on the outer box.

- · Test Cassettes: The test strips include anti-CRP antibody-coated particles and anti-CRP antibodies coated on the membrane
- Single Dilution Buffer Vials: A buffered solution containing ProClin 300 as a preservative. The Buffer is supplied ready to use.
- Capillaries
- Droppers
- Package Insert

MATERIALS NOT PROVIDED

- Specimen collection tubes
- Timer
- Centrifuge

PRECAUTIONS

- For in vitro diagnostic use only
- Do not use after the expiration date indicated on the package. Do not use the test if the foil 2. pouch is damaged. Do not reuse tests.
- 3 This kit contains products of animal origin. Certified knowledge of the origin and/or sanitary state of the animals does not completely guarantee the absence of transmissible pathogenic agents. It is therefore recommended that these products be treated as potentially infectious, and handled observing usual safety precautions (eg. do not ingest or inhale).
- 4 Avoid cross-contamination of specimens by using a new specimen collection container for each specimen obtained.
- 5 Read the entire procedure carefully prior to any testing.
- Do not eat, drink or smoke in the area where the specimens and kits are handled. Handle all specimens as if they contain infectious agents. Observe established precautions against microbiological hazards throughout the procedure and follow standard procedures for proper disposal of specimens. Wear protective clothing such as laboratory coats, disposable gloves and eve protection when specimens are assaved.
- Do not interchange or mix reagents from different lots. 7
- Humidity and temperature can adversely affect results. 8
- Used testing materials should be discarded in accordance with local regulations. 9

HAZARD and PRECAUTIONARY STATEMENTS

Refer to the SDS, available at www.meridianbioscience.com for Hazard and Precautionary Statements.

SHELF LIFE AND STORAGE

The kit should be stored at 2-30 C until the expiry date printed on the sealed pouch. The test must remain in the sealed pouch until use. DO NOT FREEZE. Care should be taken to protect the components of the kit from contamination. Do not use if there is evidence of microbial contamination or precipitation. Biological contamination of dispensing equipment, containers or reagents can lead to false results.

SPECIMEN COLLECTION AND PREPARATION

Preparation

- Before performing the test, please make sure that all components are brought to room temperature (15-30 C). Cold buffer solution or moisture condensation on the membrane can lead to invalid test results
- 2 Take a single Dilution Buffer Vial out of the kit. Label it with patient's ID. Open the screw cap

Blood Sample Collection

Collect the specimen according to standard procedures.

- Do not leave specimens at room temperature for prolonged periods. Serum and plasma specimens may be stored at 2-8 for three days. For long term storage, specimens should be kept below -20 C. Whole blood collected by venipuncture should be stored at 2-8 C if the test is to be used within two days of collection. Do not freeze whole blood specimens. Whole blood collected by fingerstick should be tested immediately. 2.
- Bring specimens to room temperature prior to testing. Frozen specimens must be completely thawed and mixed well prior to testing. Avoid repeated freezing and thawing of specimens. 3 EDTA-, citrate- or heparin-anticoagulated blood can be used. Before performing the test,
- the sample has to be diluted accordingly with the supplied buffer.

Specimen Dilution / Stability

- With the capillary tube, aspirate 10 µL blood. It is important that the capillary is filled end to end to ensure 10 µL blood.
- 2. Place the end-to-end blood-filled capillary into the dilution buffer vial. Alternatively, the 10 µL of specimen can be added directly with the micropipette into the sample Dilution Buffer
- 3. Close the vial and shake the sample vigorously for approximately 10 seconds so that sample and dilution buffer mix well. (See Figure-1)
- Let the diluted sample rest for approximately 1 minute. 4.
- The diluted specimen can then be used immediately or stored for up to eight hours. 5.



TEST PROCEDURE

Bring tests, specimens, Buffer, and/or controls to room temperature (15-30 C) before use.

- Remove the Test Cassette from the sealed pouch, and place it on a clean, level surface. Label the device with patient or control identification. For best results, the assay should be performed within one hour
- 2. Open the tube containing specimen. Transfer 3 drops of the specimen to the sample well. Start the timer.
- 3. Wait for the colored lines to appear. The result should be read at 5 minutes. Do not interpret the result after 10 minutes



INTERPRETATION OF RESULTS

POSITIVE: Three colored bands appear on the membrane. One band appears in Control region (C) and one band in the Reference region (R). Another band should appear in the test region (T).

- A test band (T) signal which is weaker than the R indicates a CRP level between 10 and 30 mg/L.
- A test band (T) signal which is close to R indicates a CRP level about 30mg/L

A test band (T) signal which is stronger than the R indicates a CRP level above 30 mg/L. NEGATIVE: Colored lines appear in both the control (C) and reference (R) regions. No colored line appears in the test line region (T). It indicates a CRP level less than 10 mg/L.

INVALID: Control band or Reference band fail to appear. Results from any test unit, which has not produced Control band or Reference band at the specified read time must be discarded. Please review the procedure and repeat with a new test. If the problem persists, discontinue using the kit immediately and contact Meridian's Technical Services Department at 800-343-3858 or your local distributor. NOTE:

- The color intensity in the test region (T) may vary depending on the concentration of CRP present in the specimen. Therefore, any shade of color in the test region should be considered positive. Please note that this is a semi-quantitative test only, and cannot determine the concentration of CRP in the specimen.
- 2. Insufficient specimen volume, incorrect operating procedure or expired tests are the most likely reasons for control band failure.

OUALITY CONTROL

This test should be performed per applicable local, state, or federal regulations or accrediting agencies.

Internal procedural controls are included in the test. Control brand and reference band appearing in the reference regions are considered internal procedural controls, confirming sufficient specimen volume and correct procedural technique.

External controls are not supplied with this kit. It is recommended that positive and negative external controls be tested as a good laboratory practice to confirm the test procedure and to verify proper test performance

If the expected control reactions are not observed, repeat the control tests as the first step in determining the root cause of the failure. If control failures are repeated please contact Meridian's Technical Services Department at 1-800-343-3858 (US) or your local distributor.

EXPECTED VALUES

TruQuick CRP has been evaluated with a leading commercial CRP EIA and correlates well.

LIMITATIONS OF THE PROCEDURE

- 1. TruQuick CRP is for in vitro diagnostic use, and should only be used for the semiquantitative detection of C - reactive protein.
- TruQuick CRP will only indicate the semiquantitative level of CRP in the specimen and 2. should not be used as the sole criteria for evaluating inflammatory conditions.
- 3. Like with all diagnostic tests, a confirmed diagnosis should only be made by a physician after all clinical and laboratory findings have been evaluated.
- 4. CRP values near the cut-off level (10 mg/L) and reference line (R: 30 mg/L) should be reported with caution as with all semiguantitative assays there exists some level of variation. Therefore, a T line with slightly higher intensity than R can also represent a value slightly below 30 mg/L. A repeat test or further quantitative test is recommended in such cases.
- High concentrations of CRP may produce a dose hook effect, resulting in incorrect 5. interpretation of CRP levels. High dose hook effect has not been observed with this test up to 2,000 mg/L of CRP.

SPECIFIC PERFORMANCE CHARACTERISTICS Sensitivity and Specificity

TruQuick CRP has been evaluated with a leading commercial CRP EIA test using clinical specimens. The results show that the sensitivity of TruQuick CRP is 98.8% and the specificity is 98.7% relative to the leading EIA test.

Method		EIA		Total Beault
TruQuick CRP	Results	Positive	Negative	Total Result
	Positive	79	4	83
	Negative	1	296	297
Total Result		80	300	380

Sensitivity: 79/80 = 98.8% (95% CI*: 95.6%~100%);

Specificity: 296/300 = 98.7% (95% CI*: 96.6%~99.6%);

Correlation: (79+296)/(79+1+4+296) = 98,7%(95% CI*; 97,0%~99,6%). *Confidence Intervals

REPRODUCIBILITY

Inter-Assay Precision

Negative, CRP low positive, CRP middle positive and CRP high positive samples were run in 10 replciates individually on 10 separate days using the same lot of TruQuick CRP. The expected results were obtained with all replicates in serum and plasma. Intra-Assav Precision

Negative, CRP low positive, CRP middle positive and CRP high positive samples were run in replicates of 10 in three separate lots of product. The expected results were obtained with all replicates in serum and plasma

CROSSREACTIVITY

TruQuick CRP was tested with HBsAg, HBsAb, HBeAg, HBeAb, HBcAb, anti-HIV, Syphilis, HAMA, RF, Mononucleosis, anti-CMV, anti-Rubella, anti-Toxoplasmosis, anti-H. pylori positive specimens. The samples showed no crossreactivity.

TESTS FOR INTERFERING SUBSTANCES

The following potentially interfering substances were added to CRP negative and positive specimens

Acetaminophen 20 mg/dL	Caffeine 20 mg/dL				
Acetylsalicylic acid 20 mg/dL	Gentisic acid 20 mg/dL				
Ascorbic acid 20 mg/dL	Albumin 10,500 mg/dL				
Creatine 200 mg/dL	Hemoglobin 1,000 mg/dL				
Bilirubin 1,000 mg/dL	Oxalic acid 600 mg/dL				
Cholesterol 800 mg/dL	Triglycerides 1,600 mg/dL				
None of the substances interfered in the assay at the concentration tested.					

REFERENCES

- Morley JJ, Kushner. Serum C-reactive protein levels in disease. In: Kushner I, Volanakis JE, Gewurz H. eds. C-reactive protein and the plasma protein response to tissue injury. Ann. NY Acad. Sci. 1982:389:406-417
- 2 Peltola HO. C-reactive protein for rapid monitoring of infections of the central nervous system. Lancet: 1982;980-983.
- 3. Macy EM, Hayes TE, Tracy RP. Variability in the measurement of C-reactive protein in healthy subjects: implications for reference intervals and epidemiological applications. Clin. Chem. 1997;43,52-58.

For technical assistance, call Technical Support Services at 800-343-3858 between the hours of 8AM and 6PM, USA Eastern Standard Time. To place an order, call Customer Service Department at 800-543-1980.

эy	y guide to symbols						
	Σ	Use By	CONTROL +	Positive control			
	LOT	Batch Code	CONTROL -	Negative control			
	IVD	In vitro diagnostic medical device	EC REP	Authorized Representative in the European Community			
	CE	This product fulfills the requirements of Directive 98/79/EC on in vitro diagnostic medical devices	SMP PREP DIL SPE	Sample Preparation Apparatus containing Sample Diluent			
	REF	Catalogue number	\otimes	Do not freeze			
	[]i	Consult Instructions for Use	BUF RXN	Reaction Buffer			
	***	Manufacturer	ľ	For IVD Performance Evaluation Only			
	Σ	Contains sufficient for <n> tests</n>	SOLN STOP	Stopping Solution			
	X	Temperature limitation	CONJ ENZ	Enzyme Conjugate			
	SN	Serial number	CONTROL	Assay Control			
	TEST	Test Device	REAG	Reagent			
	~~	Date of manufacture	BUF WASH	Wash Buffer			
	BUF	Buffer	\wedge	Warning			
	CONJ	Conjugate	DIL SPE	Specimen Diluent (or Sample Diluent)			
	SUBS	Substrate	BUF WASH 20X	Wash Buffer Concentration: 20X			
	RUO	Research Use Only	DET REAG	Detection Reagent			
	IUO	Investigational Use Only	R _∗ Only	Prescription Use Only			
	8	Do not use if package is damaged					

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SYMBOL USAGE You may see one or more of these symbols on the labeling/packaging of this product:



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